



## CURRENT METHODS FOR THE DIAGNOSIS OF PERTUSSIS INFECTIONS

### Speakers

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Dr. Pawloski obtained her PhD in Genetics from the University of Georgia. She began her work at CDC as an ORISE fellow in the Pertussis and Diphtheria Laboratory in 2005 to assist in the development of serological diagnostic assays. Since then, she has collaborated with FDA to develop and evaluate the IgG Anti-PT ELISA kit that has proven to be a very useful diagnostic tool for pertussis in public health laboratories, surveillance, and outbreak response. Dr. Pawloski published the analytical validation of the IgG Anti-PT ELISA kit and co-authored several other publications on pertussis and diphtheria in peer-reviewed journals.

**Lauren G. Pittenger, PhD, MBA**, Senior Consultant, Booz Allen Hamilton, Atlanta, GA

Dr. Pittenger has worked on a range of projects including laboratory capacity modeling for the influenza group, production and procurement support for the Laboratory Response Branch, Project management for the Bacterial Rapid Response and Advanced Technology laboratory, and QMS implementations for several groups at CDC. She is an adjunct Professor of Microbiology at Georgia Perimeter College.

Previously, she worked as Laboratory Manager/Grad Student for the USDA- ARS, in Athens GA. While at USDA she developed a microarray for studying *Campylobacter jejuni*, developed DNA sequencing capabilities for the research station, and managed the lab.

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Dr. Tatti has over twenty-five years of experience in bacteriology, virology and molecular biology. In 2005, she joined the Pertussis and Diphtheria Laboratory (PDL). As a member of PDL at CDC, she has worked on molecular diagnostics of pertussis and developed a multiplex PCR assay for the diagnosis of pertussis. She has authored and co-authored over 40 peer-reviewed manuscripts, reports and book chapters, several of which involve the laboratory diagnosis of pertussis.

### Objectives

At the conclusion of this program, participants will be able to:

- Discuss current testing methods for pertussis diagnosis: real-time PCR, serology and culture.
- Describe the two new assays developed by CDC and how they could add value to your current testing algorithm.
- Develop a greater understanding for the need to move towards standardized testing.

Continuing education credit is no longer available for this program

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For more information, visit [www.aphl.org/courses](http://www.aphl.org/courses)

# Current Methods for Diagnosis of Pertussis Infections

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**Pertussis and Diphtheria Laboratory**  
Meningitis and Vaccine Preventable Diseases Branch  
Division of Bacterial Diseases  
National Center for Immunization and Respiratory Diseases  
Centers for Disease Control and Prevention

Atlanta, GA  
January 21, 2010

## Presented by

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## Objectives

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- Understand current testing methods for pertussis diagnosis and implications in public health
  - Culture
  - Real-time PCR
  - Serology
- Become familiar with the two new assays developed by CDC and how they could add value to your current testing algorithm
  - Multiplex PCR
  - IgG anti-PT ELISA
- Realize the importance of complementary testing
- Develop a greater understanding for the need to move towards standardized testing

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## Outline

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- Background
  - Epidemiology
  - Diagnostics
    - Culture
    - Real-Time PCR
    - Serology
  - Conclusions
  - Future Directions
- Lauren Pittenger
- Kathleen Tatti
- Lucia Pawloski

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## Pertussis Background and Culture

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## Pertussis Background

- Severe, debilitating cough illness ("100 day cough")
- Highest morbidity and mortality rates in infants
- Despite high vaccine coverage, remains a public health problem
- Clinical diagnosis and laboratory confirmation can be challenging
- Outbreaks regularly occur

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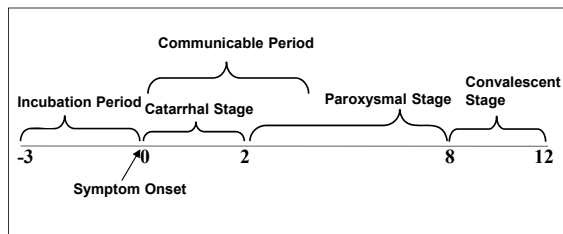
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## Stages of Disease in Weeks




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## Pertussis CSTE Case Definition

- Clinical case definition
  - Cough  $\geq 2$  weeks and at least one symptom: paroxysms, whoop, posttussive vomiting
- Case classification
  - Confirmed cases
    - Culture positive
    - Clinical case and PCR positive
    - Clinical case and epi-linked to confirmed case
  - Probable case
    - Only meets the clinical case definition

\* Serology not yet included in the case definition/classification

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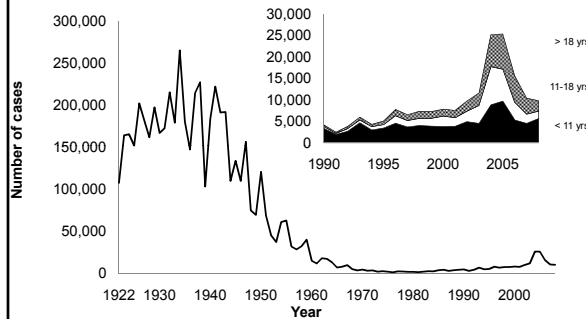
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## Reported Pertussis Cases United States 1922-2008

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\*SOURCE: CDC, National Notifiable Diseases Surveillance System and Supplemental Pertussis Surveillance System and 1922-1949, passive reports to the Public Health Service

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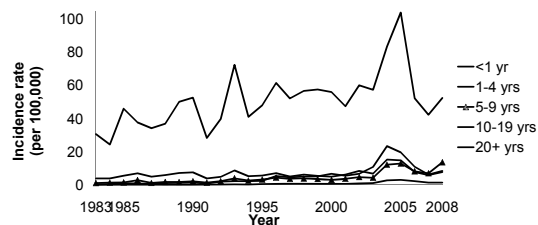
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## Reported Pertussis Incidence by Age Group, US 1983-2008

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Source: CDC, National Notifiable Diseases Surveillance System and Supplemental Pertussis Surveillance System and passive reports to the Public Health Service

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## Diagnostic Needs

### Clinical vs. Public Health

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- Clinical setting
  - Optimizes sensitivity
  - Rapid turnover
- Public health setting
  - Optimizes specificity
  - Confirmation of etiology
  - Prevention and control measures

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## Outbreaks that Exhibit the Need<sup>9</sup> for Complementary Testing

- Hospital: Winter 2006
- School: Winter 2007
- Community: Summer 2009




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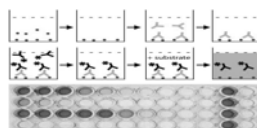
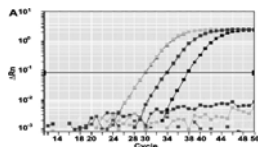
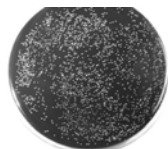
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## Pertussis Diagnostics

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- Culture
- Real-Time PCR (R-PCR)
- Serology




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Handout

## Diagnostic Tests for Pertussis

Test	Sensitivity	Specificity	Optimal Timing	Advantages	Disadvantages
Culture	12 – 60%	~ 100%	< 2 weeks post-cough onset	Very specific (100%)	Low sensitivity; 7-10 day delay between specimen collection and diagnosis
PCR	70 – 99%	86 – 100%	< 4 weeks post-cough onset	Rapid test; more sensitive than culture; organisms do not need to be viable; positive post-antibiotics	No FDA approved tests or standardization; potential for false positives; DNA contamination is problematic
Paired* Sera	90 – 92%	72 – 100%	At symptom onset and 4-6 weeks later	Effective indication of mounting antibody titers	Late diagnosis; no FDA approved tests or standardization
Single* Sera	36 – 76%	99%	At least 2 weeks post-cough onset; ideally >4 weeks post-cough	Useful for late diagnosis post antibiotics	No FDA approved test or standardization; possibly confounded by recent vaccination; diagnostic cut-offs not validated

\*Not part of CDC/CSTE case definition (Exception: MA single point ELISA assay)

\*\* Sensitivity and specificity values obtained from Wendelboe and Van Rie, 2006.

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## Specimen Collection



- Specimen type will impact ability to isolate bacterium
  - Nasopharyngeal (NP) aspirates yield similar or higher rates of recovery than NP swabs (rayon or polyester)
  - Throat and anterior nasal swabs yield unacceptably low rates of recovery

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## After Specimen Collection

- Plate immediately or place into Regan-Lowe transport medium
- Dispensing and plating should be completed within 24 hours of specimen collection
- Specimen can be used for both culture and PCR




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## Culture

- “Gold Standard”
  - Essential for public health labs
- 100% specific, but low sensitivity
  - Most sensitive within first two weeks after cough onset
- Highest yield
  - Young patients
  - Unvaccinated patients
  - Patients early in cough illness prior to antimicrobials
- Incubation time 4-10 days
- Although specific collection methods, transport, media and growth conditions are needed, culture is not difficult



Gram stain of *B. pertussis*

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## Methods for Culturing Pertussis

- Regan-Lowe or Bordet-Gengou media
- Inoculate on media with and without antibiotics
- 35-36°C incubation with high humidity
- Ensure plates do not dry out
  - Plastic bags
  - Canisters
  - Pan of water
- Check plates every day




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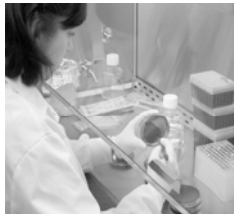
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## Growth Characteristics

- Bordet-Gengou (BG)
  - Small colony size
  - Appearance similar to mercury droplets
  - Colonies appear hemolytic
- Regan-Lowe (RL)
  - Small colony size
  - Glistening, cut glass appearance




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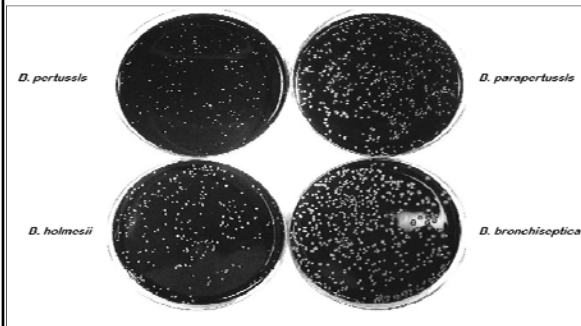
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## *Bordetella* on RL Medium

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## Other *Bordetella* spp.

- *B. parapertussis*
  - Colonies typically appear within two-three days
  - On RL agar the colonies will appear greyish
  - On BG agar colonies have a brown pigmentation
- *B. holmesii*
  - Colonies look similar to *B. pertussis*
  - Growth is inhibited by cephalixin
- *B. bronchiseptica*
  - Large colonies
  - Appear after one day
  - On RL agar the colonies will have a slight brown coloration

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## Public Health Impact of Pertussis Culture

17

- Particularly important if an outbreak is suspected
- Isolation of the bacterium confirms pertussis
  - Other respiratory pathogens often cause similar clinical symptoms
  - Co-infection with other pathogens does occur
  - Colony morphology helps to identify other species of *Bordetella*
- Necessary for antimicrobial susceptibility testing and molecular typing

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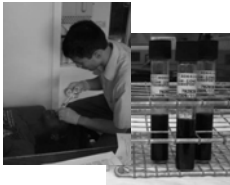
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## Culture is Feasible School Outbreak 2007

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- Proper transport medium allowed excellent recovery of *B. pertussis* when direct plating could not be performed
  - Culture was performed at CDC
  - Overnight shipment was not available




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## Pertussis Real-Time PCR

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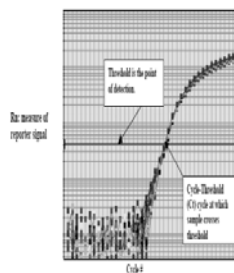
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## R-PCR Assay-IS481

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- Present in three *Bordetella* species
  - 50 to >200 copies in *B. pertussis*
  - 8 to 10 copies in *B. holmesii*
  - One copy in *B. bronchiseptica* (host specific)
- High Ct value could indicate
  - Positive test result
  - False positive
  - Positive result of a *Bordetella* species other than *B. pertussis*

Real-Time PCR Amplification Plot




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Available online at [www.sciencedirect.com](http://www.sciencedirect.com)

ScienceDirect

Diagnostic Microbiology and Infectious Disease 61 (2009) 264–272

[www.elsevier.com/locate/diagmicrobio](http://www.elsevier.com/locate/diagmicrobio)

DIAGNOSTIC  
MICROBIOLOGY  
AND INFECTIOUS  
DISEASE

# Development and evaluation of dual-target real-time polymerase chain reaction assays to detect *Bordetella* spp.<sup>☆</sup>

Kathleen M. Tatti<sup>a,1</sup>, Kai-Hui Wu<sup>1</sup>, Maria Lucia Tondella, Pamela K. Cassidy,  
Margaret M. Cortese, Patricia P. Wilkins, Gary N. Sanden

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Received 1 November 2007; accepted 27 February 2008



**MMWR**<sup>TM</sup>

Morbidity and Mortality Weekly Report

Weekly

August 24, 2007 / Vol. 56 / No. 33

## Outbreaks of Respiratory Illness Mistakenly Attributed to Pertussis — New Hampshire, Massachusetts, and Tennessee, 2004–2006

<http://www.cdc.gov/mmwr/preview/mmwrhtml/mm5633a1.htm>

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## MMWR Article

- Outbreaks were attributed to pertussis based only on local single-target PCR (IS481 assay)
- 2 specimens were positive for *B. holmesii* by PCR
- Only one specimen was positive by CDC two-target assay
- PCR results were not confirmed by the two-target assay at CDC
- Limitations of relying solely on a single-target PCR to confirm pertussis outbreaks
- Emphasizes importance of
  - Diagnostics validation
  - PCR result interpretation
  - Standardization of PCR assays

23

### Benefit of Dual Target-Hospital Outbreak 2006

AB 7000 N=111	<i>ptxS1</i> +	<i>ptxS1</i> -
IS481: Ct<35	1	2
IS481: 35≤Ct<40	0	22
IS481 -	0	86

MMWR, Outbreaks of respiratory illness mistakenly attributed to pertussis, 2004-2006. 2007;56:837-842

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### Decontamination



- Throughout this process
  - Keep your lab coat on and wear gloves
- Four step process
  - 10% bleach
  - 70% ethanol
  - DNA AWAY ®
  - UV exposure for a minimum of one hour

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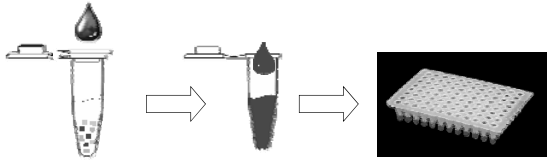
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### Multiplex R-PCR Assay for *Bordetella* spp.

■ □ ○ IS481 primers/probe set FAM- *B. pertussis*/*B. holmesii*  
■ □ ○ hIS1001 primers/probe set Quasar- *B. holmesii*  
■ □ ○ pIS1001 primers/probe set HEX- *B. parapertussis*



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## Multi-target R-PCR Allows for Speciation

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Species	ptxS1	Multiplex		
		IS481	hIS1001	pIS1001
<i>B. pertussis</i>	+	+	-	-
<i>B. paraptussis</i>	+	-	-	+
<i>B. pertussis</i> and <i>B. paraptussis</i>	+	+	-	+
<i>B. holmesii</i>	-	+	+	-

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Handout

## Multi-target R-PCR Approach

- The multiplex assay utilizes three target sequences
  - IS481
  - hIS1001 specific for *B. holmesii* (3-5 copies/cell)
  - pIS1001 targets *B. paraptussis* (20-23 copies/cell)
- The ptxS1 targets the gene for the S1 subunit of pertussis toxin
  - Single copy in *B. pertussis* and *B. paraptussis*
- Additional assay targets human *maseP*

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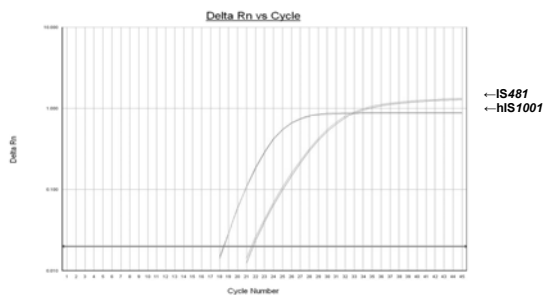
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## Clinical Specimen with *B. holmesii* Hospital Outbreak 2006

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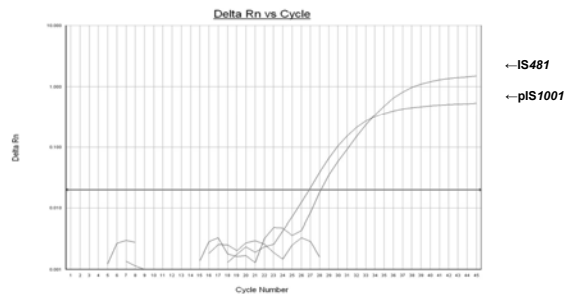
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## Clinical Specimen with *B. pertussis* and *B. parapertussis*

28



## Analytical Sensitivity and Clinical Relevance

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Genomic equivalents	<i>Bordetella pertussis</i> ptxS1 Ct values	<i>Bordetella pertussis</i> IS481 Ct values
1000	29	19
100	32	23
10	35	26
1	39	30
0.1	Negative	33

## Analytical Sensitivity and Clinical Relevance

30

Genomic equivalents	<i>Bordetella pertussis</i> ptxS1 Ct values	<i>Bordetella pertussis</i> IS481 Ct values	<i>Bordetella parapertussis</i> IS1001 Ct values	<i>Bordetella holmesii</i> hIS1001 Ct values
1000	29	19	22	25
100	32	23	26	29
10	35	26	30	33
1	39	30	33	38
0.1	Negative	33	36	Negative

CDC R-PCR  
Pertussis Outbreak Algorithm

	IS481+ (Ct<35)	IS481+ (35≤Ct<40)	IS481- (Ct≥40)
ptxS1+ (Ct<40)	<i>B. pertussis</i>	<i>B. pertussis</i>	<i>B. paraptussis</i> <sup>1</sup>
ptxS1- (Ct≥40)	<i>B. holmesii</i> <sup>2</sup>	Indeterminate	Negative

<sup>1</sup> Confirmed by pIS1001 target

<sup>2</sup> Confirmed by hIS1001 target

R-PCR Interpretation Criteria

- Interpret R-PCR results along with the clinical symptoms and epidemiological information
- Determine the clinically relevant cut-off value for all targets
- Use indeterminate as a result for multi-copy target for IS481, not ptxS1

Whooping Challenges  
Respiratory Outbreak Investigation in Colorado, U.S.

CDC investigates mysterious pertussis cases in Durango


August 20th, 2009, 8:25 am · Post a Comment · posted by Brian Newsome

A team from the U.S. Centers for Disease Control and Prevention has flown from Atlanta to Durango to investigate a spike in pertussis cases with unusual symptoms, according to the health department there.

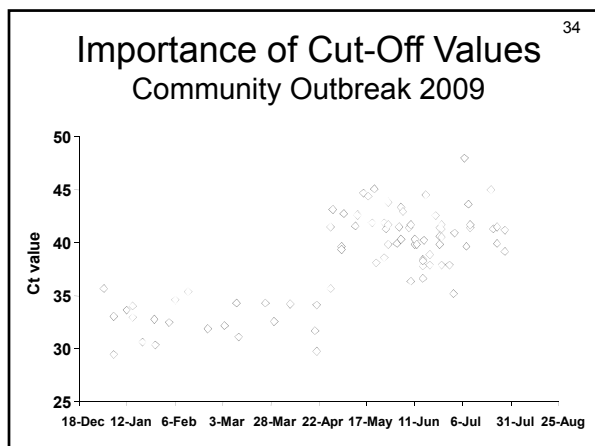
The San Juan Basin Health Department says the spike in whooping cough, in La Plata County since 2007, even though the Southwest Colorado region has not reported any cases since 2000.

Many of the cases are mild and don't have the classic whooping cough, the health department says. The unusually high number of cases prompted the Colorado Department of Health to send a team to investigate.

The four-person team is interviewing people to help find a cause.



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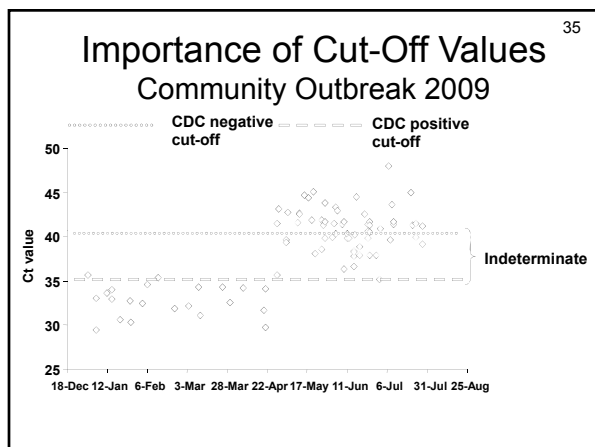
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### Addressing the Myths of a Multi-target Approach

Concern	Resolution
• Will reduce sensitivity	• Data has shown that there is no reduction in sensitivity
• Multiplex is more expensive	• Initial cost may be higher • Fewer re-tests
• It will take longer to generate data	• Assay does not take longer to run than a single target assay
• It is a technically difficult assay to perform	• No more difficult than a single target PCR
• New equipment will have to be purchased	• Maybe, some older platforms can't accommodate multiplex

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## Pertussis Serology

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## Pertussis Serology

- Originally designed for vaccine evaluation
- Routinely used for diagnosis in other countries
- Included as part of Massachusetts' case definition
- Useful for confirming diagnosis, especially during outbreaks where culture is not performed
- Can be used later in disease

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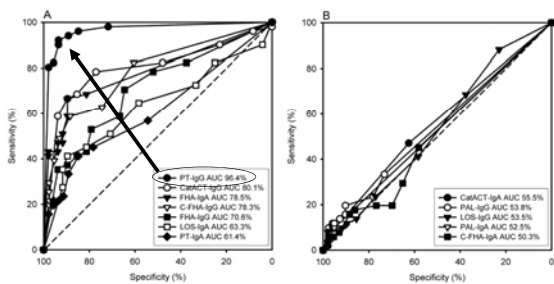
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## IgG Against Pertussis Toxin (PT) is the Most Specific and Sensitive Target<sup>39</sup>




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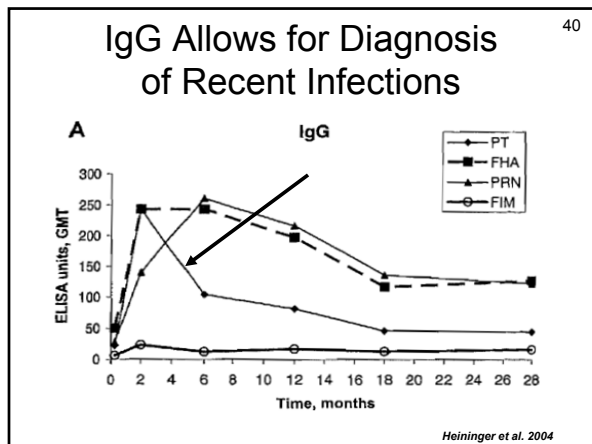
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### CDC/FDA IgG Anti-PT ELISA Kit

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- If present, specific IgG antibodies in the serum will bind to the PT adsorbed to the wells
- The concentration of the PT IgG antibodies is directly proportional to the intensity of the color

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### Easy to Use

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- 6 ready-to-use standards
- 3 ready-to-use controls
  - 49, 94 EU/mL and negative
- Standard curve can be generated
- Simple assay / production process
  - Single dilution of serum
  - Minimal reagent preparation
  - Incubation at room temperature
  - Calibrated to reference sera
  - Monoclonal antibody conjugate

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Handout

## IgG Anti-PT ELISA Kit Protocol

- Day 1
  - Coat PT onto microtiter wells
  - Incubate for 14-20 hours
- Day 2
  - Wash; add standards, controls, and test samples (1:100 diluted)
  - Incubate 2 hours at RT
  - Wash and add the conjugate antibody
  - Incubate 2 hours at RT
  - Wash and add the substrate
  - Incubate 10 minutes at RT
  - Add stop solution
  - Read at 450 nm



43

CLINICAL AND VACCINE IMMUNOLOGY, Dec. 2009, p. 1761-1768  
1556-6811/09/\$12.00 DOI:10.1128/CVI.00248-09  
Copyright © 2009, American Society for Microbiology. All Rights Reserved.

Vol. 16, No. 12

### Development and Analytical Validation of an Immunoassay for Quantifying Serum Anti-Pertussis Toxin Antibodies Resulting from *Bordetella pertussis* Infection<sup>7</sup>

Sandra L. Menzies,<sup>1,\*</sup> Vijay Kadwad,<sup>1,4</sup> Lucia C. Pawloski,<sup>2</sup> Tsai-Lien Lin,<sup>3</sup> Andrew L. Baughman,<sup>2</sup> Monte Martin,<sup>2</sup> Maria Lucia C. Tondella,<sup>2</sup> Bruce D. Meade,<sup>1,5</sup> and the Pertussis Assay Working Group

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Received 19 June 2009/Returned for modification 28 August 2009/Accepted 19 October 2009

## Addressing the Myths of Serodiagnosis

44

Concern	Resolution
• Recent vaccination may confound diagnosis	• Post-vaccination antibody levels do not interfere with diagnosis
• Paired sera is necessary	• Single serum point taken at the optimal time is sufficient
• Special analytical software is needed	• Assay can be qualitative
• It takes too long to perform	• Can be run in as few as 20hrs
• Publication of single strain lacking PT gene	• Isolates tested to date have PT gene

## Public Health Use of Pertussis Serology

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Outbreak	Sera	Positive	Indeterminate
Hospital 2006	39	1	5
School 2007	169	49	10
Community 2009	15	0	0

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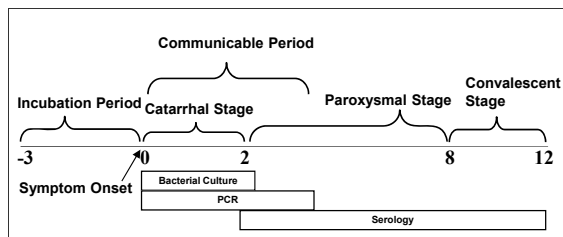
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## Optimal Timing for Diagnostic Testing

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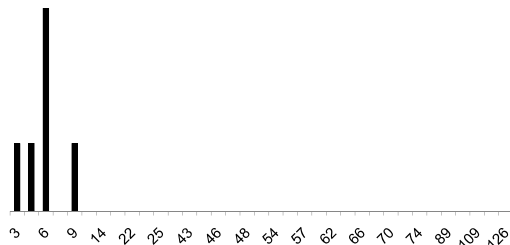
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## Test Results by Cough Duration

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School Outbreak 2007




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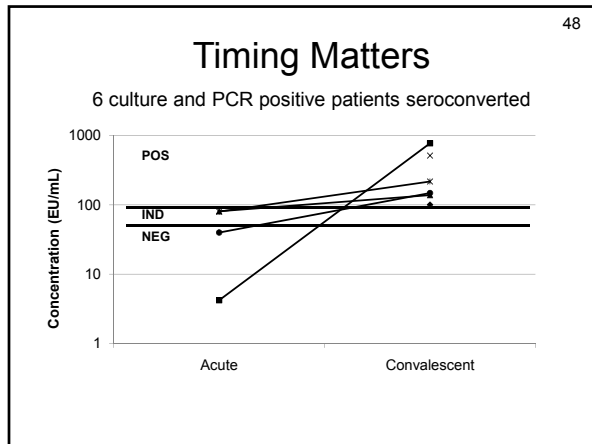
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- 49
- ### Conclusions
- No single laboratory test can stand alone for diagnosing pertussis
  - Adoption of multi-target R-PCR methods will allow for confirmation and speciation among *Bordetella* spp
  - Serology is a useful method for diagnosing pertussis especially in adults and in the later stages of the disease
  - Labs should maintain culture capabilities
  - Standardization is needed
  - Diagnosis is an important part of surveillance

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- 50
- ### CDC can Provide
- Protocols and draft SOPs for testing methods
  - Culture
    - Support and advice on techniques
  - PCR
    - Support with running on CDC validated PCR platforms: ABI 7500 and other platforms
    - Provide guidance on use of many extraction methods
  - Serology
    - Support and advice on development, implementation, and standardization of ELISA

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## Future Directions Clinical Validation Study

- Estimate the clinical sensitivity, specificity and predictive values of diagnostic tests:
  - CDC/FDA IgG anti-PT ELISA
  - CDC's combined multi-target R-PCR
- Assess clinical usefulness as related to
  - Stage of disease, age, antibiotic use and vaccination status

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## How can State and Local Labs Help?

- Help with early identification of outbreaks
  - Contact:  
Meningitis and Vaccine Preventable Diseases Branch  
404-639-3158  
Ask for the duty officer
- Send isolates to CDC
- Adopt standardized assays when available

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## Pertussis Website

**Pertussis (Whooping Cough)**

**Laboratory Information**

The CDC provides laboratory support for United States (US) health departments requesting assistance with isolation, identification, and subtyping of *Bordetella pertussis*, *B. pertussis*, *B. parapertussis*, *B. bronchiseptica*, *C. jejuni*, and *C. coli*. This laboratory serves as a reference and outbreak support for state and local health departments, research and observational investigators, non-acute laboratory and hospital personnel. In addition to serving as a resource for other laboratories, the Pertussis and C. jejuni Laboratory at CDC provides assistance with the development, evaluation, implementation, and improvement of molecular and serologic methods, techniques and techniques to enhance the diagnosis and surveillance of agents causing pertussis and dysentery.

**On This Page**

- Laboratory Information
- Reference Lab
- Pathogens Isolated at the CDC Reference and Diagnostic Laboratory
- HDT010005

**Contact Us:**

Centers for Disease Control and Prevention  
1600 Clifton Rd  
Atlanta, GA 30333

800 CDC BPDU  
1800 232-6200  
1101 (888) 232-6348  
24 Hours/7 Days 1-800-FDA-1088

<http://www.cdc.gov/pertussis/lab.html>  
(Under Development)

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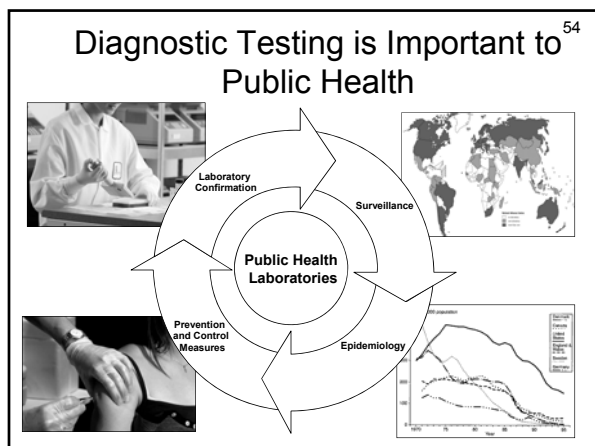
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- 55
- ## Acknowledgements
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  - APHL
    - Kathleen Breckenridge
    - Travis Jobe
    - Rosemary Humes

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## Contacts

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The findings and conclusions in this report have not been formally disseminated by the Centers for Disease Control and Prevention/the Agency for Toxic Substances and Disease Registry and should not be construed to represent any agency determination or policy.

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Handout

## References

Heininger U, Cherry JD, Stehr K. Serologic response and antibody-titer decay in adults with pertussis. Clin Infect Dis. 2004 Feb 15;38(4):591-4. Epub 2004 Jan 29.

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Wendelboe AM, Van Rie A. Diagnosis of pertussis: a historical review and recent developments. Expert Rev Mol Diagn. 2006 Nov;6(6):857-64.

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